

**Description**

HOT START DNA polymerase is an ideal enzyme for high fidelity PCR with excellent processivity. It is a novel engineered enzyme with comparable performance to Pyrococcus DNA polymerase. With unique structure HS DNA polymerase contains a recombinant synthesis enhancement domain to increase fidelity and extension speed. The antibody-mediated hot-start feature significantly inhibits non-specific amplifications at room temperature. HOT START is one of the thermostable DNA polymerases with strong 3' -5' exonuclease activity which results in its extreme high fidelity, 10-15 times higher than Taq DNA polymerase and 6 times higher than Pyrococcus furiosus DNA polymerase.

The HS PCR Kit is supplied with a 5X HF Buffer and a 2.5X GC Buffer. The 5X HF Buffer is an optimized buffer for general high fidelity amplifications while the 2.5X GC Buffer is used in the amplifications of problematic or GC-rich templates.

Contents	FS-T-2131-200
Hot Start DNA Polymerase (2000U/mL)	200 RNXS
5X HF PCR Buffer	2x 1 mL
2.5X GC PCR Buffer	4X 1 mL
dNTPs (10 mM each)	200 µL

**Thermal Inactivation:** No

**Product End:** Blunt end

**Standard Protocol**

- It is recommended to prepare all reaction components on ice, and then quickly transfer the reaction system to a thermocycler preheated to 98°C. It is recommended to prepare all reaction.
- All components should be mixed and collected at the bottom of a tube with a quick spin before use. Add Hot Start 2X Master Mix with Dye at the end to prevent primer degradation by its strong 3' -5' exonuclease activity. Note: The Hot Start DNA polymerase requires special reaction conditions different from other polymerase protocols. Please refer to the recommended reaction conditions below for the better amplification yields.

**Recommended Protocol**

On ice, prepare each of following master mixes, combine, and place in heated (to 95°C) thermal cyler:

**5X HF(High Fidelity) Buffer ReactionSystem**

Component	25 µl reaction	50 µl reaction	Final Conc.
Hot Start DNA Polymerase (2000 U/mL)	0.5 ul	1 ul	2U/50ul
5X HF PCR Buffer	5 ul	10 ul	1 X
dNTP mix (10 mM each)	0.5 ul	1 ul	0.2mM
DNA Template	variable	variable	<300ng
Forward Primer (10µM)	0.5 ul	1 ul	0.2uM
Reverse Primer(10µM)	0.5 ul	1 ul	0.2uM
Distilled water	to 25 ul	to 50ul	N/A

**2.5 X GC BUFFER Reaction System**

Component	25 µl reaction	50 µl reaction	Final Conc.
Hot Start DNA Polymerase (2000 U/mL)	0.5 ul	1 ul	2U/50ul
2,5X GC PCR Buffer	10 ul	20 ul	1x
dNTP mix (10 mM each)	0.5ul	1ul	0.2mM
DNA Template	Variable	variable	< 300 ng
Forward Primer (10µM)	0.5 ul	1 ul	0.2uM
Reverse Primer (10µM)	0.5 ul	1 ul	0.2uM
Distilled water	to 25 ul	to 50ul	N/A

**General Cycling Conditions:**

Step	Temp (°C)	Time	Cycle
Initial Denaturation	98	45 sec.	1
Denature	98	10 sec.	25 ~35
Anneal	55~65	20 ~ 30 sec.	
Extend	72	10 ~ 30 sec. s/kb*	
Final Extension	72	5 min.	1
Hold	4-12		1

\*Note: Properly extending the extension time can improve the amplification yield. For complex amplification templates, such as genomic DNA, it is recommended to extend at a speed of 60 s/kb, and more recommended conditions please refer to the basic principles of PCR below.

**PCR Principles**

**1. Template**

High-quality purified DNA templates are important to high-fidelity PCR reactions. The recommended DNA template amounts with different complexity are listed Below (For a 50 µL reaction):

DNA	IMPUT Amount
Plants, animals and human gDNA	10 ng-300 ng
E.coli, lambda gDNA	10 ng-100 ng
Plasmid DNA	1 pg-10 ng

Note: If the DNA template is obtained from a cDNA synthesis reaction, the template volume should be less than 10% of the total reaction volume. If long fragments are amplified, the amount of template input should be increased appropriately.

**2. Primers**

Oligonucleotide primers are typically 20-40 nucleotides in length with a GC content of 40-60%. Primers can be designed and analyzed using software such as Primer 3. The final concentration of each primer in the PCR reaction system should be in the range of 0.1-1 µM.

**3. Enhancer**

The Enhancer solution is an optional component to increase the amplification efficiency for problematic templates, such as GC-rich sequence or genes with strong secondary structure. Note: Since the enhancer is included in the 2.5X GC Buffer, additional enhancer is not recommended with the use of 2.5X GC Buffer. Excess amount of enhancer may be inhibitory.

**4. Buffer** The HS PCR Kit contains a 5X HF Buffer and a 2.5X GC Buffer. The 5X HF Buffer is designed for general high fidelity PCR amplification, and the 2.5X GC Buffer is optimized for the amplifications of GC-rich templates.

#### **5. Denaturation**

98°C pre-denaturation for 45 s can fully denature most DNA templates. In the case of high complexity DNA templates, the pre-denaturation time should be extended up to 3 minutes for fully denaturation. Generally, the recommended denaturation condition for low-complexity DNA templates is 98°C, 5-10 s.

#### **6. Annealing**

The annealing temperature of HS DNA polymerase is usually higher than other PCR polymerases. Generally, primers longer than 20 nt are annealed at (lower primer  $T_m+3$ )°C for 10-30 s; when the primers are shorter than 20 nt, an annealing temperature equivalent to the lower primer  $T_m$  should be used. When using a new primer set for PCR reaction, we recommend a gradient PCR to determine the optimal annealing temperature. In a two-step amplification protocol, the annealing temperature should be set to the extension temperature.

#### **7. Extension**

The recommended extension temperature is 72°C. The extension time depends on the length and complexity of the amplicon. For the low-complexity amplicons (plasmid DNA), the extension condition is 10-30 s /kb. For high-complexity amplicons, such as genomic DNA, it is recommended to increase the extension time to 1 min /kb. In some cases, the extension time for cDNA templates should be less than 1 min /kb.

#### **8. Cycles**

To obtain enough yield of PCR products, 25-35 cycles are recommended.

#### **9. PCR Products**

HS DNA polymerase produces blunt-end PCR products, which might be directly used in the sequential blunt-end cloning.

#### **Storage:**

Upon receipt, store all components at -20°C.